

Use of High-dimensional and Spatial Immune Profiling to Explore Sotigalimab (CD40 Agonist) Activation of Antigen Presenting Cells and T Cells in the Tumor Microenvironment in Patients with Esophageal/Gastroesophageal Junction Cancer

Maira Soto¹, Erin L. Filbert¹, Hai Yang^{2,3}, Li Zhang^{2,3,4,6}, Stephanie Starzinski^{2,3}, Alec Starzinski^{2,3}, Alexander Cheung^{2,3}, Tony Li⁵, Frank J. Hsu¹, Andrew Ko^{2,3,6}, Lawrence Fong^{2,3,6}, Bridget P. Keenan^{2,3,6}

¹Apexigen, Inc, San Carlos, CA, USA; ²Cancer Immunotherapy Program, University of California, San Francisco, CA, USA; ³Helen Diller Family Comprehensive Cancer Center, University of California, San Francisco, CA, USA;

Abstract # 450

⁴Department of Epidemiology and Biostatistics, University of California, San Francisco, CA, USA; ⁵Department of Genome Sciences, University of Washington, Seattle, WA, USA; ⁶Division of Hematology/Oncology, University of California, San Francisco, CA, USA

UCSF

University of California
San Francisco

Bridget.Keenan@ucsf.edu

Background

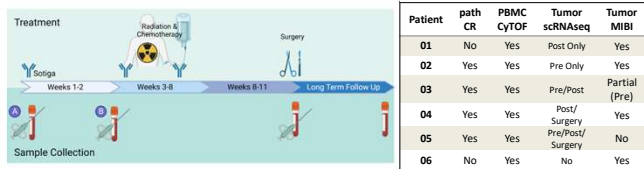
Neoadjuvant chemoradiation (CRT) followed by surgical resection is standard of care for patients with locally advanced esophageal/gastroesophageal junction (E/GJ) cancer. A pathologic complete response (pCR) at surgery is associated with improved survival outcomes. Sotigalimab (sotiga) is a potent CD40 agonist mAb capable of inducing and expanding anti-tumor immune responses.

A recently reported phase II clinical trial of sotiga combined with CRT in E/GJ cancer patients showed increased pCR rates that compared favorably to historical data¹. Here, deep immune profiling was performed on samples from the circulation and tumor microenvironment (TME) from a subset of patients to gain insight into the mechanism of sotiga.

Registered clinicaltrials.gov #NCT03165994

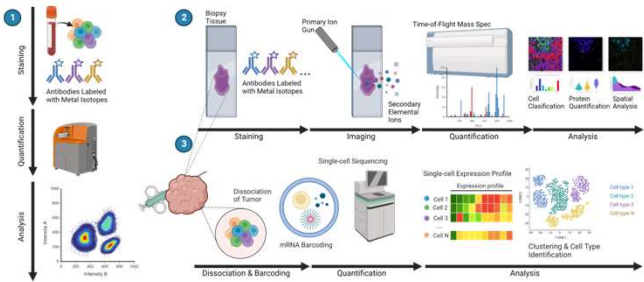
References: 1. Ko A, et al. A Multicenter Phase 2 Study of Sotigalimab (CD40 Agonist) in Combination with Neoadjuvant Chemoradiation for Resectable Esophageal and Gastroesophageal Junction (GEJ) Cancers. *ESMO Annual Conference*; 2022. 2. Spitzer, M, et al. An Interactive Reference Framework for Modeling a Dynamic Immune System. *Science*; 2015. 3. Wolf, FA, et al. SCANPY: large-scale single-cell gene expression data analysis. *Genome Biology*; 2018.

Study Protocol and Biopsy Schedule



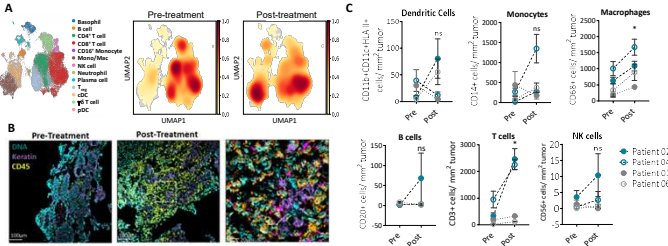
Blood samples (PBMC) were collected before and after treatment, and after surgery (N=6). Paired tumor biopsies were collected before (A) and after (B) sotiga treatment (N=4). When available, fresh tumor was collected from surgical specimens.

Analytical Methods



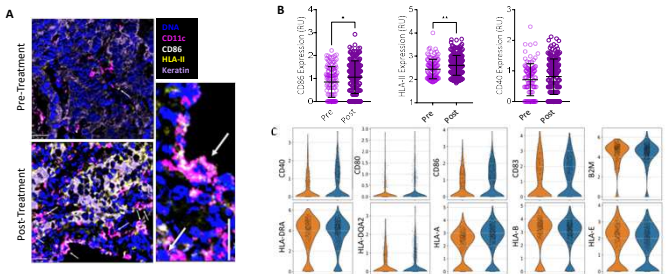
- PBMCs were isolated from whole blood, stained and analyzed by mass cytometry (CyTOF). Scaffold was used to analyze CyTOF data.²
- Tumor samples from biopsies were analyzed by IonPath using Multiplexed Ion Beam Imaging Technology (MIBI).
- Tumor samples were also dissociated and single cell RNA sequencing (scRNAseq) and T cell receptor (TCR) sequencing was performed using the 10X Genomics platform. Data was analyzed using SCANPY.³

Sotiga Induces Immune Cell Infiltration in the TME



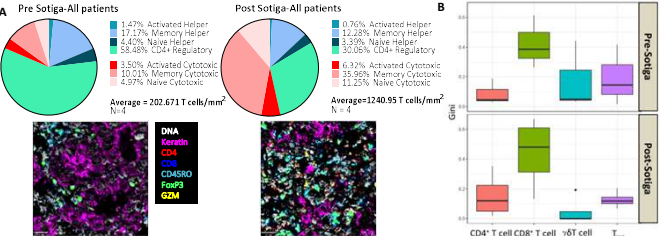
A. Uniform Manifold Approximation and Projection (UMAP) plot demonstrating all immune cells from tumor biopsies with cell type over-laid (left) or by density heatmap showing relative abundance of cell type by pre/post-treatment, demonstrating altered immune cell composition with sotiga. **B.** Staining of tumor tissue pre and post-treatment by MIBI showing increased immune infiltrate post-sotiga. **C.** Quantification of cell type using MIBI; patients that achieved pCR \bullet or did not \circ .

Infiltrating Myeloid Cells Display Inflammatory Markers



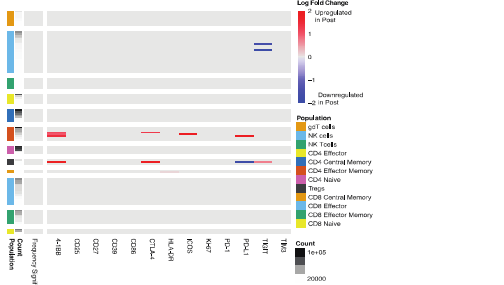
A. Images from MIBI showing increased CD11c⁺CD86⁺HLA-II⁺ myeloid cells post-sotiga. **B.** CD86 and HLA-II were significantly increased in expression on dendritic cells post-sotiga. **C.** Expression of activation markers on dendritic cells pre vs post-sotiga by scRNAseq. All transcripts shown were significantly increased post-sotiga.

Sotiga Increased Infiltration of CD8⁺ T Cells and Decreased CD4⁺ Regulatory T Cells in TME



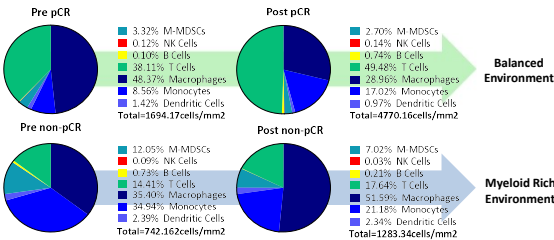
A. T cell composition pre and post-sotiga by MIBI. **B.** Gini co-efficient for T cell sub-types pre and post-sotiga, demonstrating a trend towards higher clonality of intra-tumoral CD8⁺ T cells and lower clonality of Tregs post-sotiga.

Sotiga Activates Circulating NK and T cells



Heatmap demonstrating the frequency of immune clusters and expression of activation and inhibitory markers on T and NK cells in circulation of patients with E/GJ cancer pre and post-sotiga, as analyzed using CyTOF. There were no differences in the frequency of clusters of T and NK cells, but there was increased expression of 4-1BB, CTLA-4, ICOS on T cells and up- or down-regulation of PD-L1 and TIGIT on different NK and T cell clusters.

Higher T cell Density in TME at Baseline is Associated with Tumor Response



Immune composition pre and post-sotiga in tumors that had a pCR following sotiga + CRT compared to tumors that did not have a pCR.

Conclusions

Sotiga induced dramatic changes in the tumor microenvironment and converted "cold" tumor to "hot" by increasing the frequency of activated T cells and antigen presenting cells (APCs) and decreasing frequency of T_{reg}. In circulation, treatment with sotiga activated NK and T cells. A distinct signature of T cell infiltration in baseline tumor biopsies was observed in patients who achieved a pCR versus those who did not, potentially identifying patients that may benefit from this novel treatment strategy. This is the first demonstration that single-agent sotiga can induce significant inflammatory responses in the TME. The conversion of "cold" tumors to "hot", a key MOA of sotiga, is the foundation of immunotherapy.

