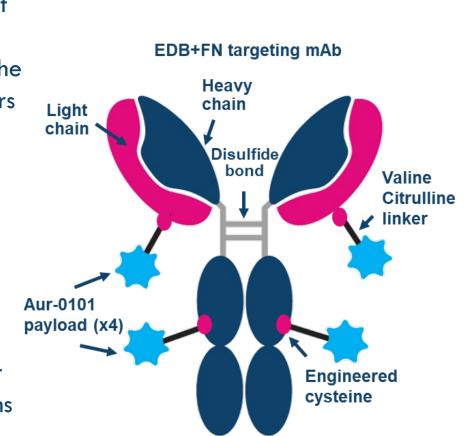
Histological biomarker analysis of nonclinical and baseline tumor samples from the phase 1 dose escalation study using micvotabart pelidotin (MICVO) in advanced solid tumors

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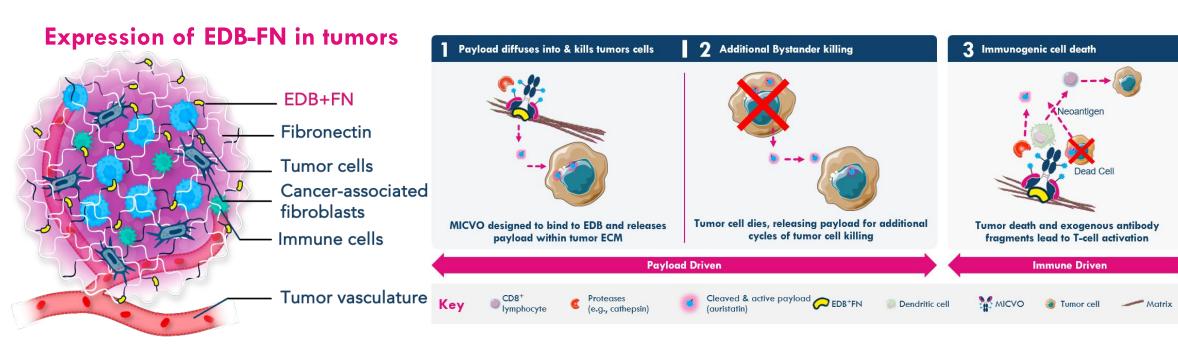
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BACKGROUND

- Micvotabart pelidotin (MICVO, aka PYX-201) is a first-in-concept antibody-drug conjugate (ADC) targeting extradomain-B of fibronectin (EDB+FN), a non-cellular structural component within the tumor extracellular matrix (ECM) that is highly expressed in tumors compared to normal adult tissues.¹
- MICVO is composed of a fully human IgG1 monoclonal antibody conjugated to an optimized Auristatin-0101 payload via a cleavable linker (DAR of 4).^{2,3}
- MICVO is designed to achieve anti-tumor activity via three mechanisms of action: 1) the cytotoxic, cell-permeable Auristatin-0101 payload directly kills tumor cells through disruption of microtubule formation, 2) the payload promotes additional tumor cell killing via the bystander effect, and 3) release of neoantigens from dying tumor cells induces immunogenic cell death.



Mechanism of Action for MICVO

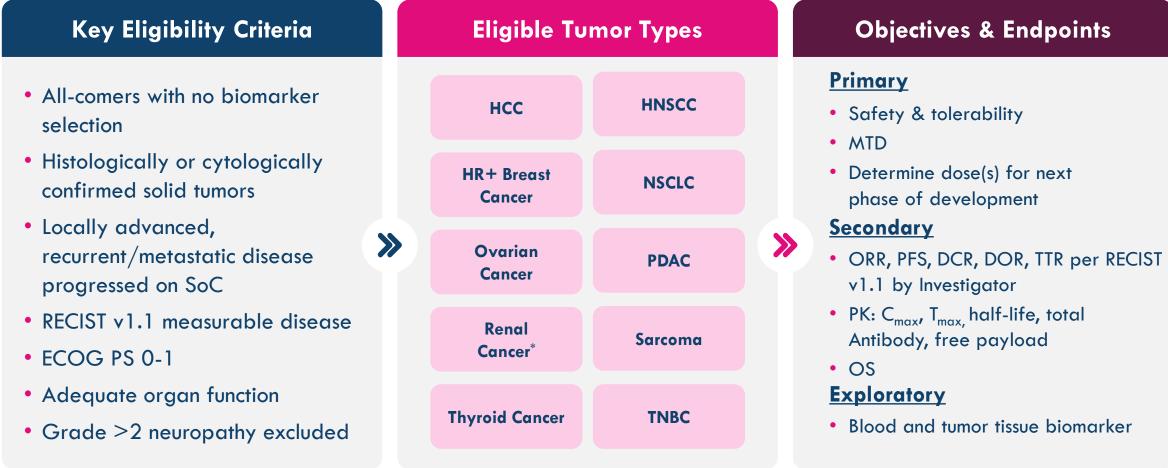


- PYX-201-101 is a first-in-human, open-label, multicenter, Phase 1 clinical study (NCT05720117) to evaluate the safety, tolerability, pharmacokinetics, pharmacodynamics, and preliminary antitumor activity of MICVO monotherapy in participants with advanced solid tumors. Study PYX-201-101 comprises two parts: Part 1 dose escalation and Part 2 dose expansion. Data from the Part 1 dose escalation is reported in ESMO Poster
- Objective: Histological biomarkers were evaluated using baseline clinical biopsy samples and nonclinical tumor samples to identify stromal features that could confer sensitivity to treatment with MICVO.

Clinical Study Design

- Treatment with MICVO IV Q3W until unacceptable toxicity or disease progression
- As of 04Oct2024, a total of 77 participants were treated with MICVO across 9 dose levels ranging from 0.3-8.0 mg/kg Q3W IV during the dose-escalation part of the study.
- Dose escalation study identified the range of potentially effective doses to be 3.6-5.4 mg/kg

Part 1 Dose Escalation

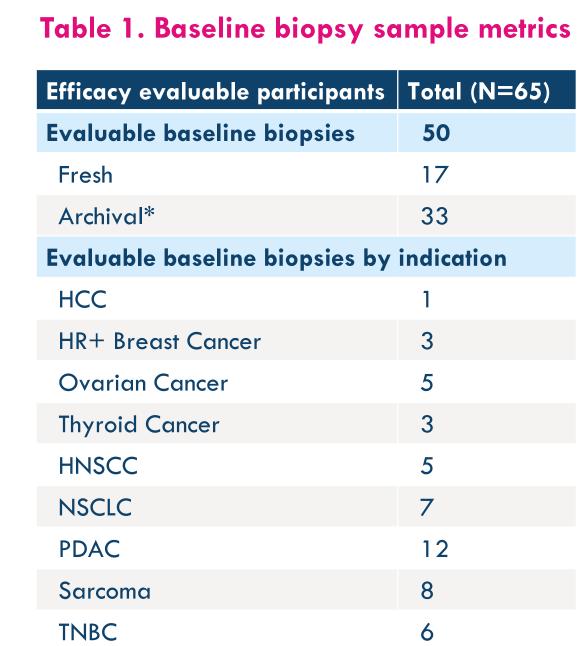


*No patient was dosed in this Phase 1 study for Renal Cancer

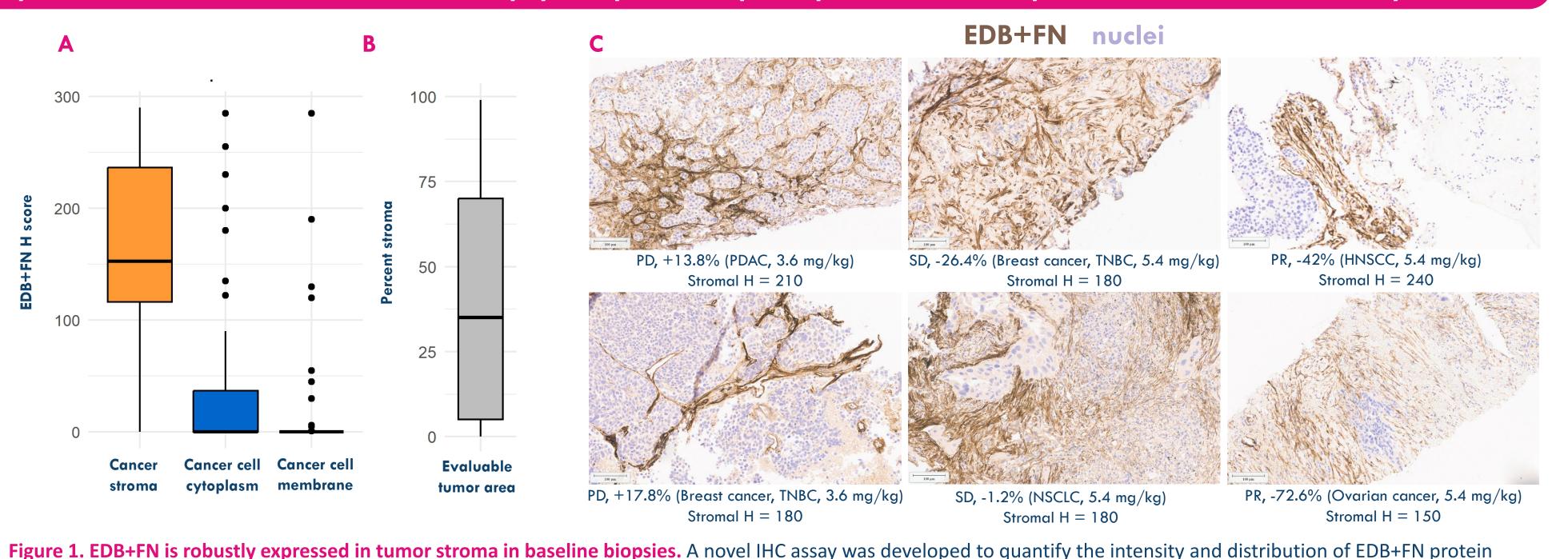
Methods

Baseline biopsies from participants in the phase 1 dose escalation study were evaluated for histological biomarkers and correlation to drug response. Tumor biopsies tested by H&E for tumor stroma and IHC for EDB+FN were scored by a board-certified pathologist for percent of tumor stroma and EDB+FN, quantified as H scores in tumor stroma, cancer cell membrane, and cancer cell cytoplasm. Scores were evaluated for correlation to clinical response. Best overall response (BOR) was per RECIST v1.1 criteria. Nonclinical human biopsy samples from tumor types of interest were evaluated for histologic features that may confer sensitivity to MICVO. Al powered digital pathology was deployed on H&E images of commercially-sourced nonclinical samples of HNSCC, NSCLC, ovarian cancer, and PDAC to annotate tissue regions and cell types and evaluate stromal architecture.

EDB+FN is robustly expressed in tumor stroma in baseline biopsy samples from participants enrolled in phase 1 dose escalation study

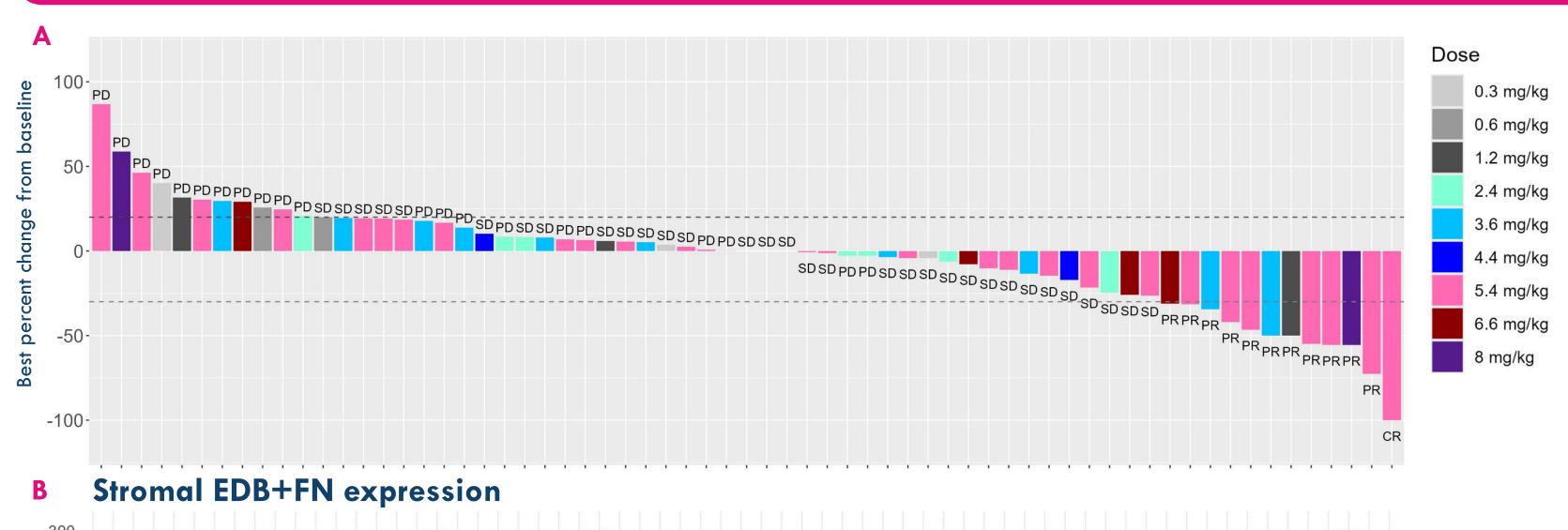


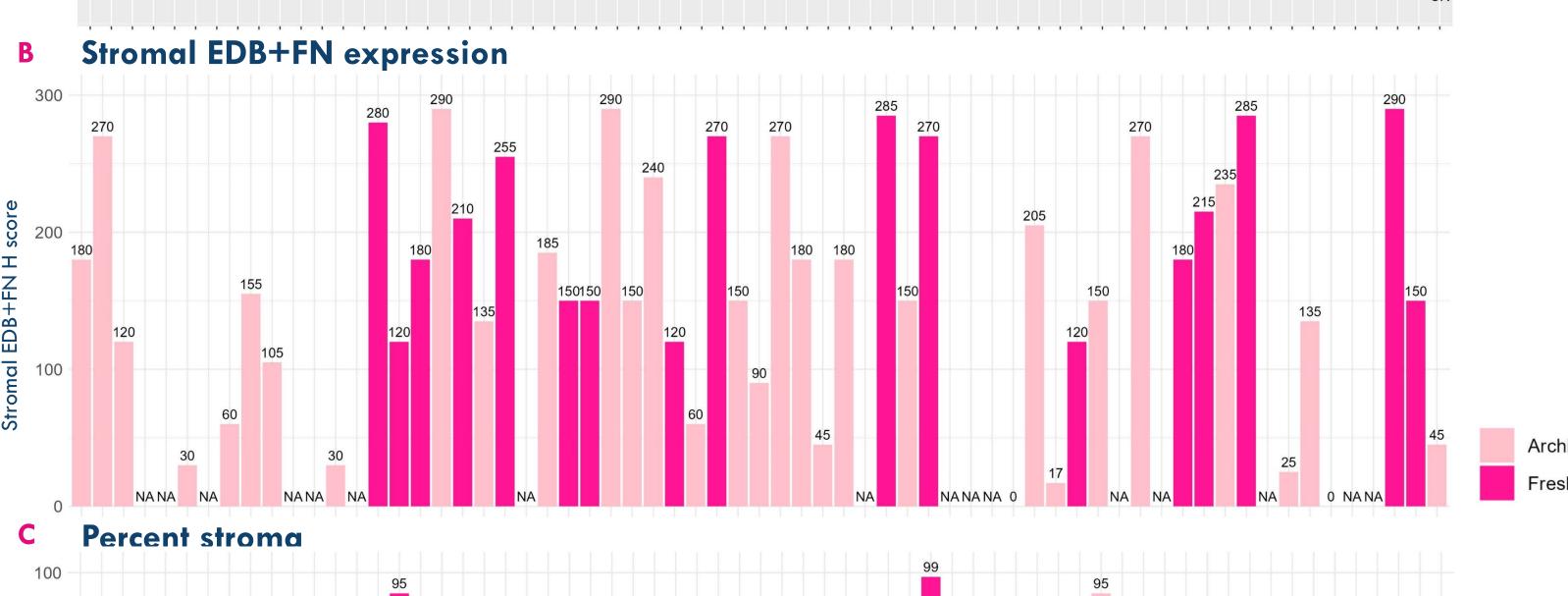




expression in the tumor stroma, cancer cell membrane, and cancer cell cytoplasm, reported as EDB+FN H-scores. All evaluable baseline biopsy samples were evaluated and scored by a board-certified pathologist for EDB+FN protein expression (A) using this assay and for percent stroma (B) using H&E staining. Cancer stroma was the predominant region of EDB+FN expression (median stromal H = 150, range 0-290), with broad expression observed across indications and clinical responses (C). Values below images reflect best overall response and best percent change from baseline.

Stromal EDB+FN expression is not directly correlated with clinical response to MICVO





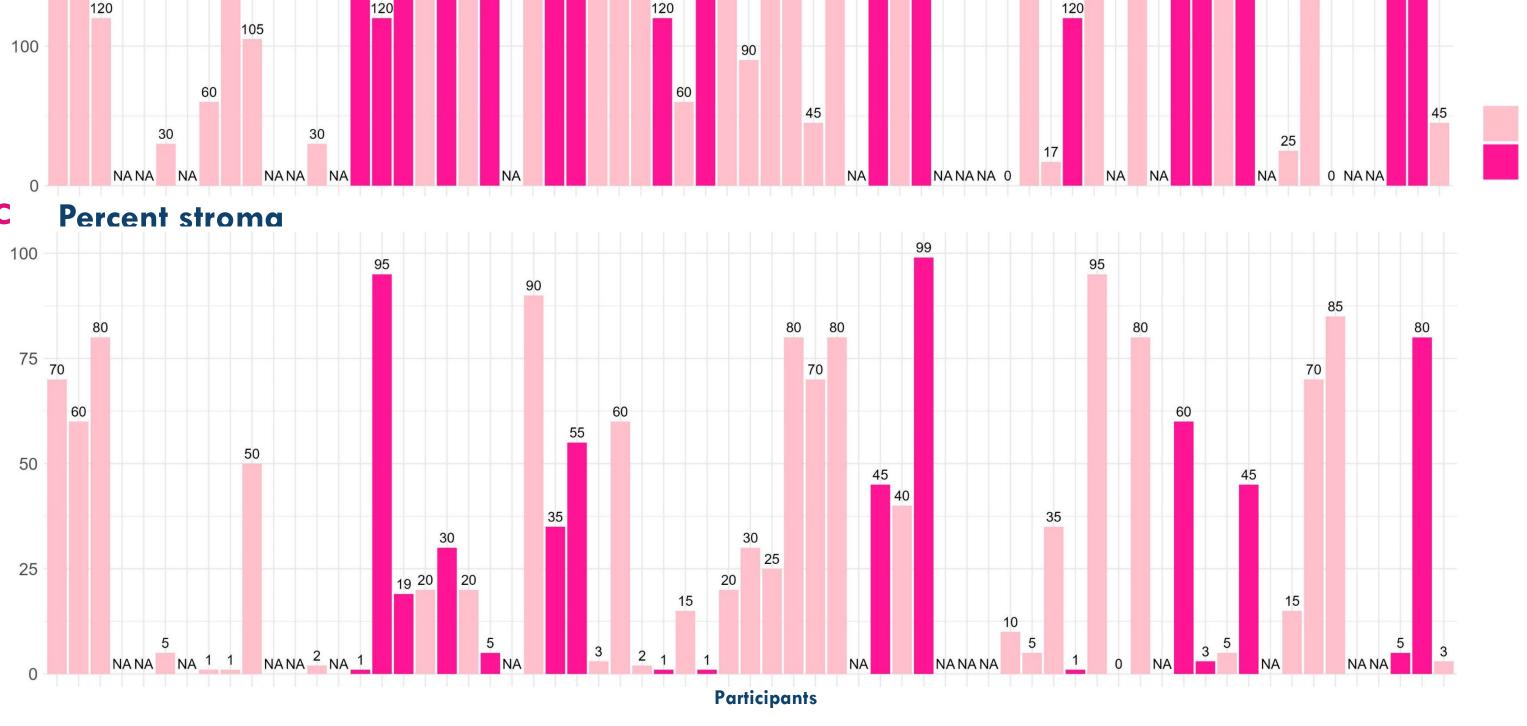
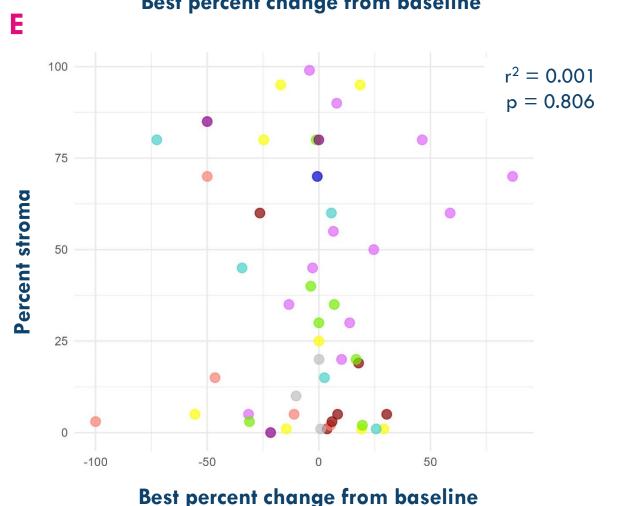


Figure 2. Clinical outcome is not directly correlated with either stromal EDB+FN expression or percent stroma. Of 67 participants with evaluable responses to MICVO per Resist v1.1 (A), 51 evaluable baseline biopsies were tested and scored by a board-certified pathologist for EDB+FN protein expression by IHC (B) and stromal density by H&E (C)*. EDB+FN protein was broadly detected in tumor stroma regardless of clinical response. Clinical response was not correlated with either EDB+FN protein expression (D) or stromal density (E), either overall or for any

In plots A-C, each bar reflects one participant, and bars are ordered by best percent sample quality. Values above bars in A reflect best overall response per RESIST v1.1. CR = complete response, PR = partial response, SD = stable disease, PD = progressive disease.

Correlation analysis p = 0.478HCC Head and Neck SCC HR+ Breast Cancer NSCLC Ovarian Cancer Pancreatic Cancer Thyroid Cancer Best percent change from baseline



Features of stromal architecture differ across indications in nonclincal samples

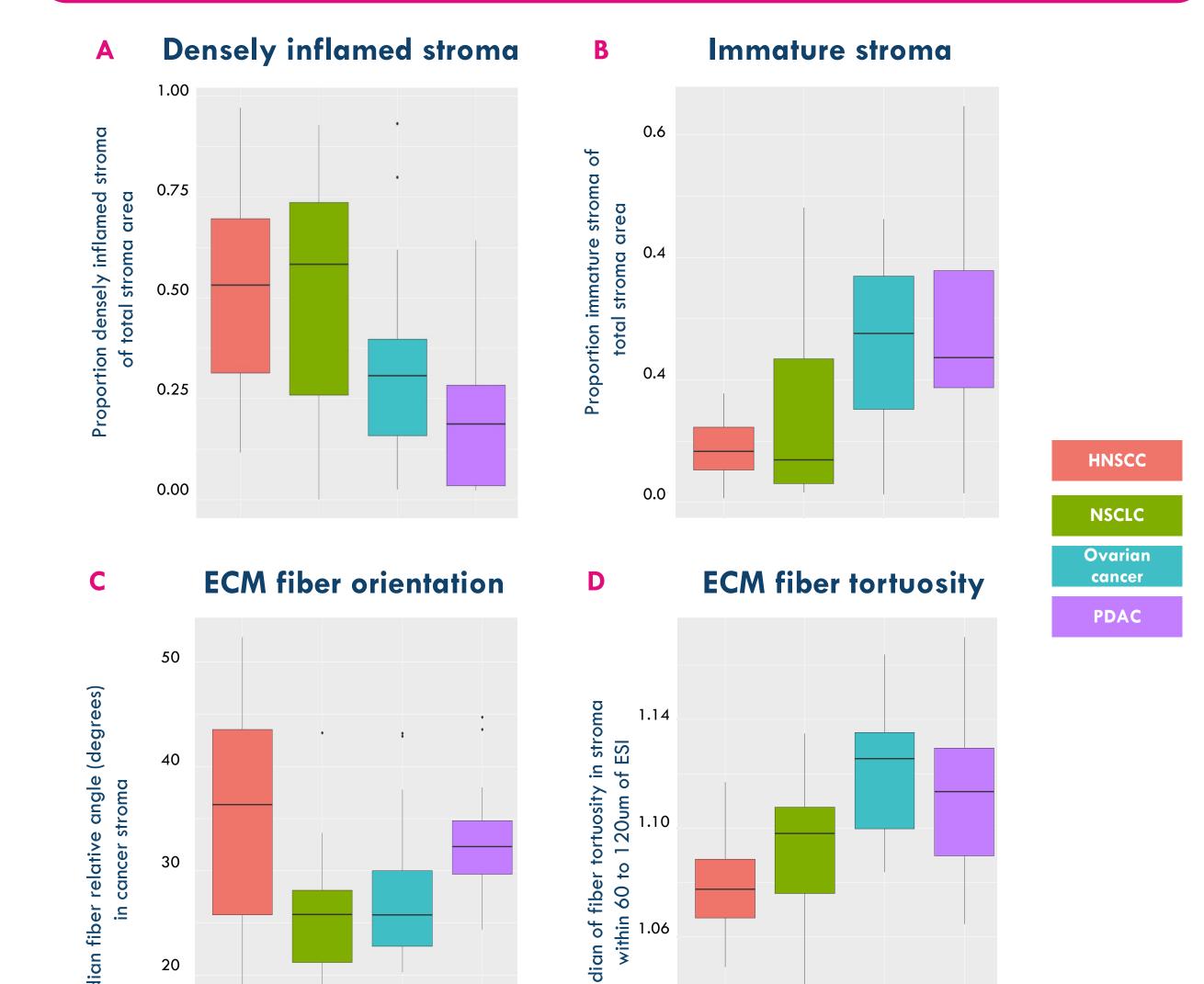


Figure 3. Digital pathology analysis of nonclinical samples revealed differences in stromal morphology between indications. To explore additional stromal features that may confer sensitivity to MICVO, Al powered digital pathology was deployed on H&E images of commercially-sourced nonclinical human tumor samples to evaluate stromal architecture across indications. Stromal subtyping analysis using PathAl PathExplore revealed that the HNSCC tumors have a higher proportion of densely inflamed stroma (A) and a lower proportion of immature stroma (B) compared to tumors from the other indications evaluated. When examined using the PathAl PathExplore fibrosis model, ECM fiber organization in HNSCC tumors showed a higher median relative fiber angle (C) and lower median fiber tortuosity (straighter fibers) 60-120 µm from the epithelial-stromal interface

(ESI) (D) compared with the fiber organization in tumors of the other indications evaluated.

SUMMARY & CONCLUSIONS

- EDB+FN is broadly expressed in tumor stroma in biopsy samples from participants in the phase 1 dose escalation study.
- In this heterogeneous dataset, the level of EDB+FN protein expression and percent stroma did not correlate with clinical response.
- In certain tumor types such as HNSCC, features of stromal architecture detected using digital pathology may contribute to sensitivity to MICVO.
- Studies with participant tumor samples correlating features besides EDB+FN such as protease expression, payload sensitivity, drug resistance mechanisms, or other histologic features such as those identified here using PathAl with sensitivity to MICVO are on-going and will be further investigated in tumor specific expansion cohorts



ACKNOWLEDGEMENTS The study was sponsored by Pyxis Oncology, Inc.. We thank all our patients and their family for participation and all research sites and CRO personnel for their support of the study. Thank you to personnel at Discovery Life Sciences and PathAl for sample testing.

DECLARATIONS OF INTEREST

Dr. Wang: Honoraria to self for lecture educational event: National Cancer Treatment Alliance; Support to self for meeting attendance: Boehringer Ingelheim (ASCO), NGM Bio (AACR); Research funding for institution only: Abbvie, Abdera Therapeutics, Accent Therap Therapeutics, Adagene, Allorion Therapeutics, Alterome Therapeutics, Apollo, Artios, Astellas Pharma, BeiGene, Bicycle Therapeutics, BioNTech SE, Biostar, Blueprint Medicines, BMS GmbH & Co. KG, Boehringer Ingelheim, C4 Therapeutics, Celgene/Bristol-Myers Squibb, Circle Pharma, Compass

Therapeutics, Compugen, Cullinan Oncology, Conjupro Biotherapeutics, D3 Bio, Daiichi Sankyo/UCB Japan, Day One Bio, Denmab, Georgiamune, GlaxoSmithKline, Halda Therapeutics, Hotspot Therapeutics, IgM

NGM Biopharmaceuticals, NiKang, Novartis, Nurix, Olema Oncology, OnCusp Therapeutics, Relay Therapeutics, Revolution Medicines, Sanofi, Step Pharma, Syndax, Systimmune, Tango Therapeutics, Vividion Therapeutics, Xencor, Zai Lab, Zymeworks

Biosciences, Immunitas, Immunogen, Incyte, ITeos Therapeutics, Janssen, Jazz Pharmaceuticals, Kineta, Klus Pharma, Medikine, Medikine, Medikine, Medikine, Medikine, Medikine, Medikine/Menarini, Merck KGaA, Mersana, Moderna Therapeutics, Immunogen, Incyte, ITeos Therapeutics, Janssen, Jazz Pharmaceuticals, Kineta, Klus Pharma, Medikine, Medikine

For additional questions on the study, please contact Marsha Crochiere at mcrochiere@pyxisoncology.com